

New frontiers in synthetic biology for spaceflight

Jonathan M. Galazka, PhD
NASA Ames Research Center
Space Biosciences Division

Funding

Funding

NASA Space Technology Mission Directorate Center Innovation Fund
NASA Advanced Exploration Systems

Team

Dr. John A. Hogan (NASA)

Dr. Asif Rahman (U. of New Mexico)

Dr. Aditya Hindupur (Wyle Labs)



NASA Ames Team Members

Dr. Michael Dougherty (Wyle Labs)
Natalie Ball (Wyle Labs)
Dr. Hiromi Kagawa (SETI)



Dr. John Hogan



Samantha Fleury & Lily Neff

WASHU Collaborators

Fuzhong Zhang Chris Bowen

Cameron Sargent Sarah Rommelfanger

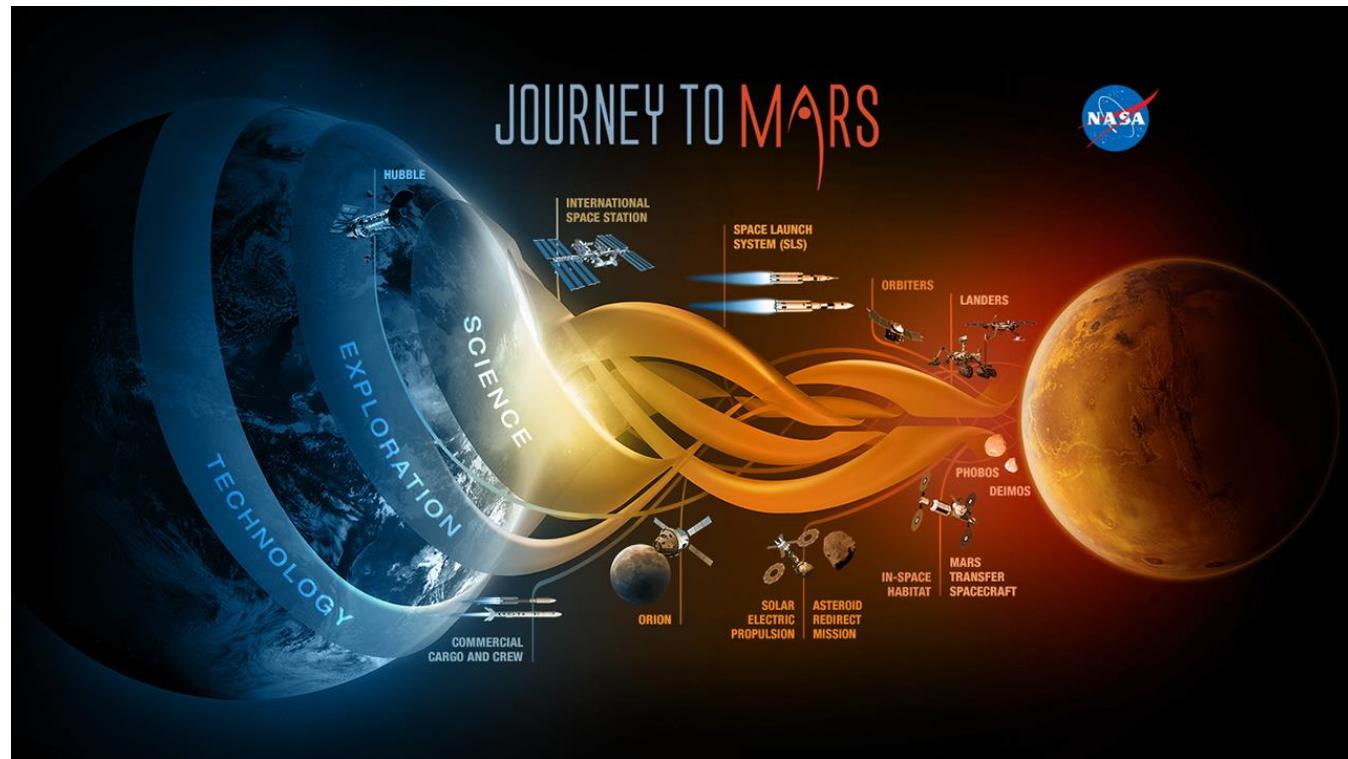
Moving to Deep Space

Challenges...

Increased radiation exposure risk

Limited opportunity for crew return (launch window occurs every 25 months)

Limited opportunity for resupply

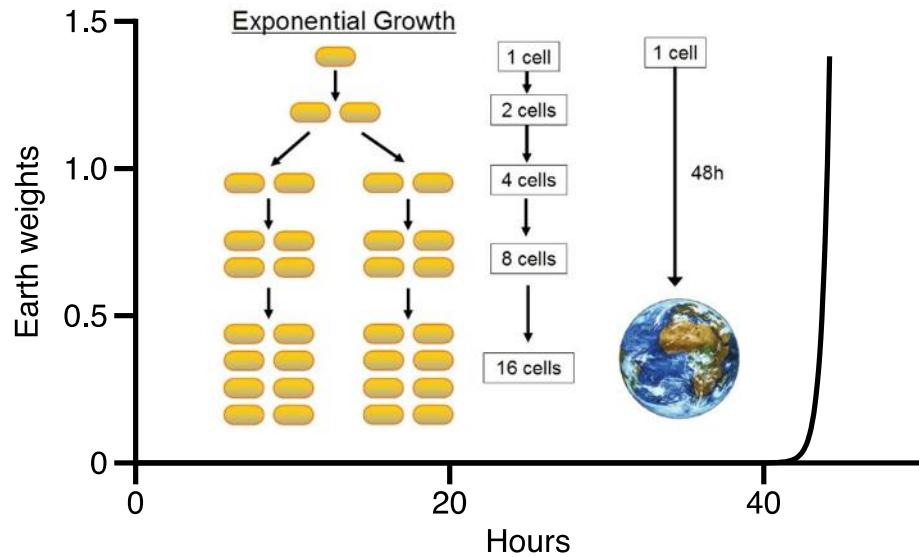


Missions to Deep Space are enabled by robust technologies that:

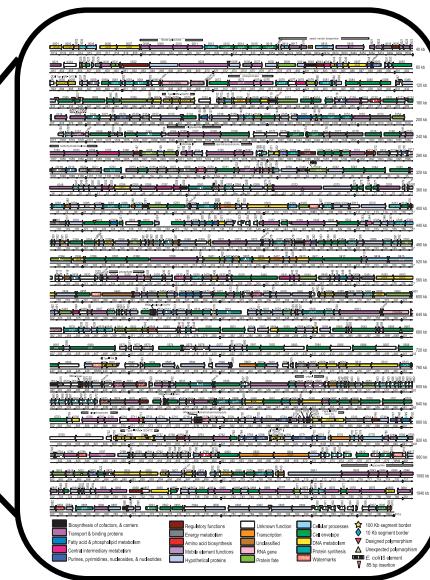
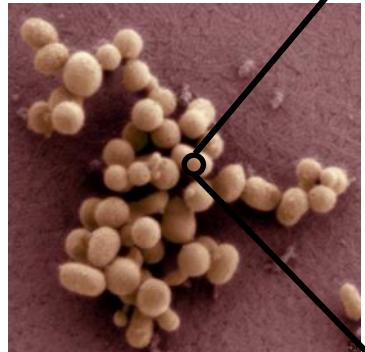
- Make crew more self-sufficient
- Allow utilization of local resources

Potential of Microbial Manufacturing in Space

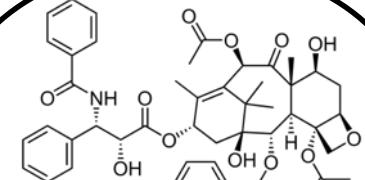
Scalable



Programmable



Self-organizing



Potential of Microbial Manufacturing in Space

Biological systems are:

- Scalable
- Programmable
- Precise (pure isomers)
- The only route of production in some cases (protein therapeutics)
- Low T° and pressure
- Regenerable



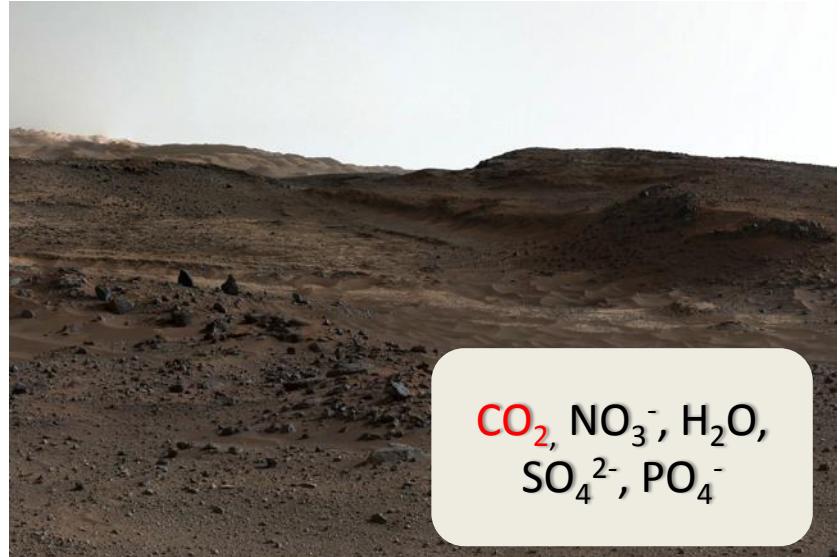
Credits: NASA

Barriers to Microbial Manufacturing in Space

Space technologies must be...

- Robust
- Simple to operate
- Stable during storage
- Compatible with available resources and infrastructure

We will have this



Mount Sharp, Mars. Credits: NASA.

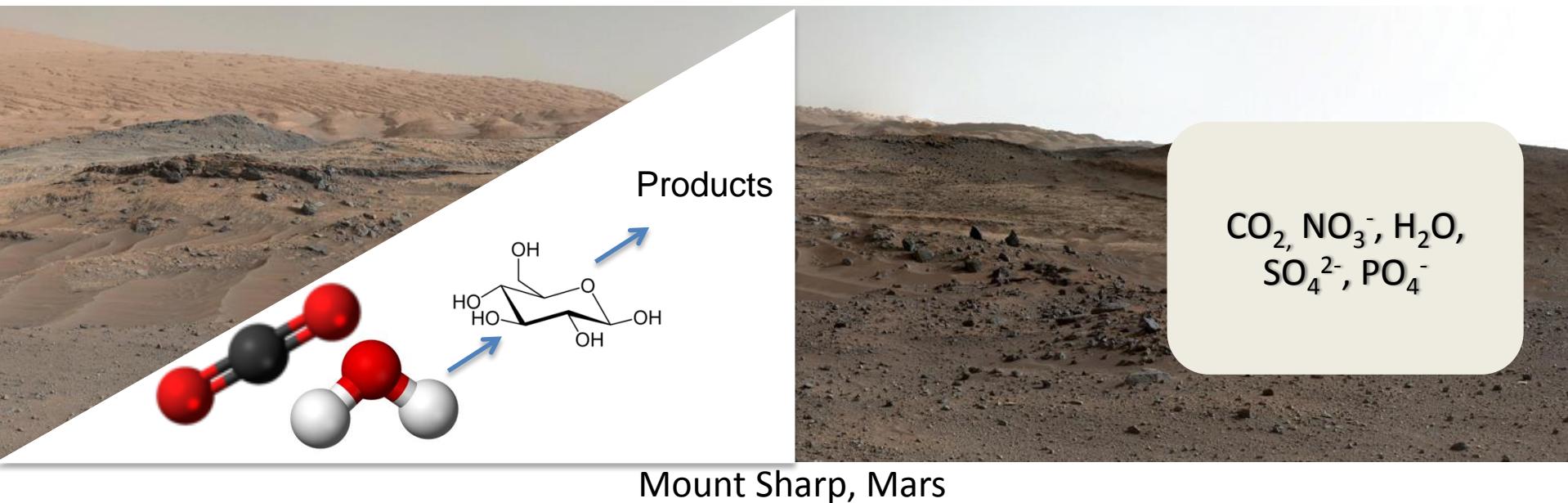
Not this



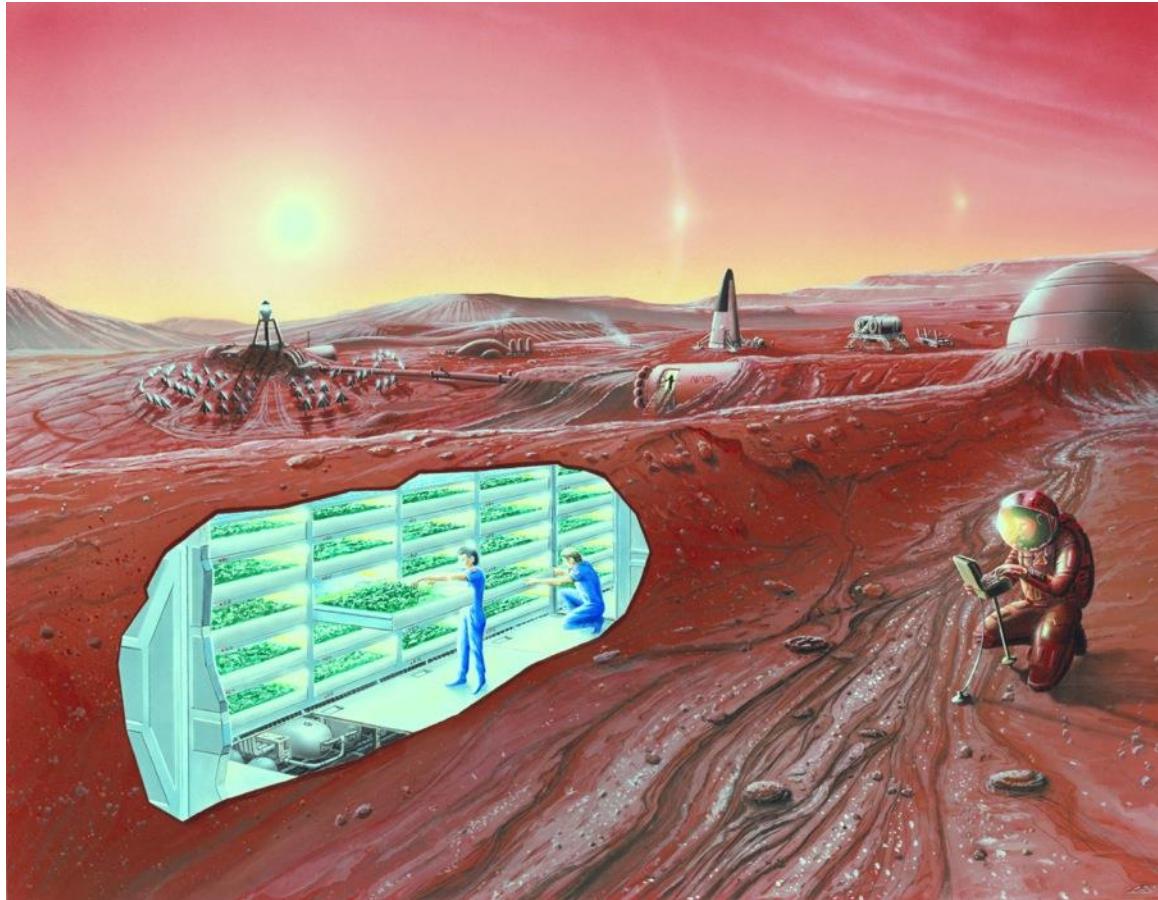
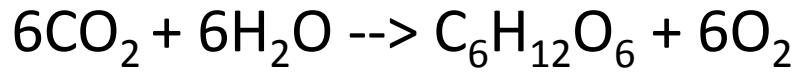
YPD:

0.3% yeast extract
1% peptone
1% glucose

Material conversion



Photoautotrophs

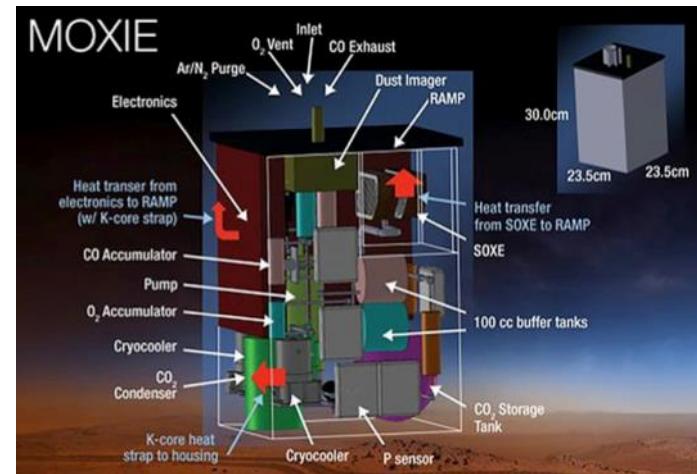
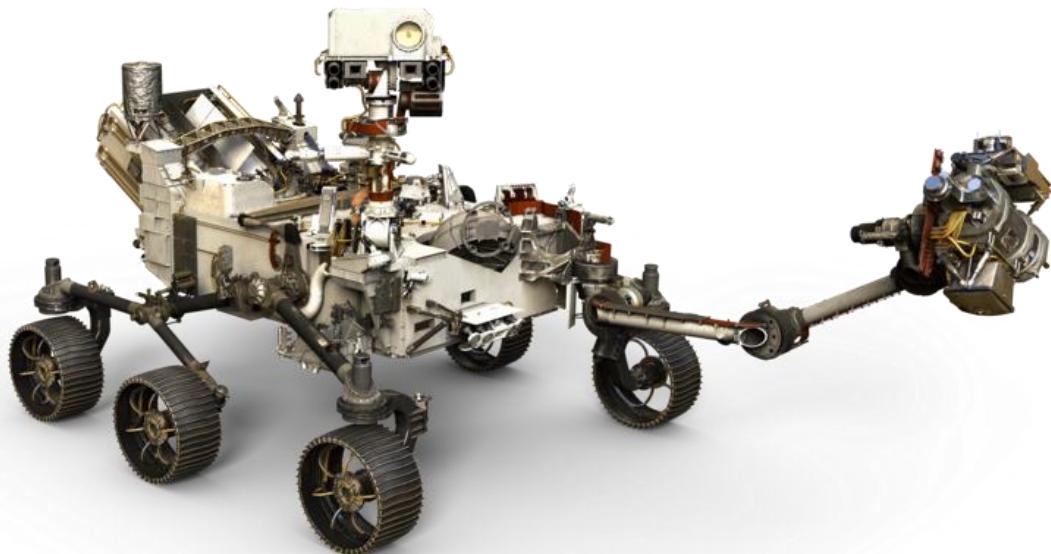


Credits: NASA

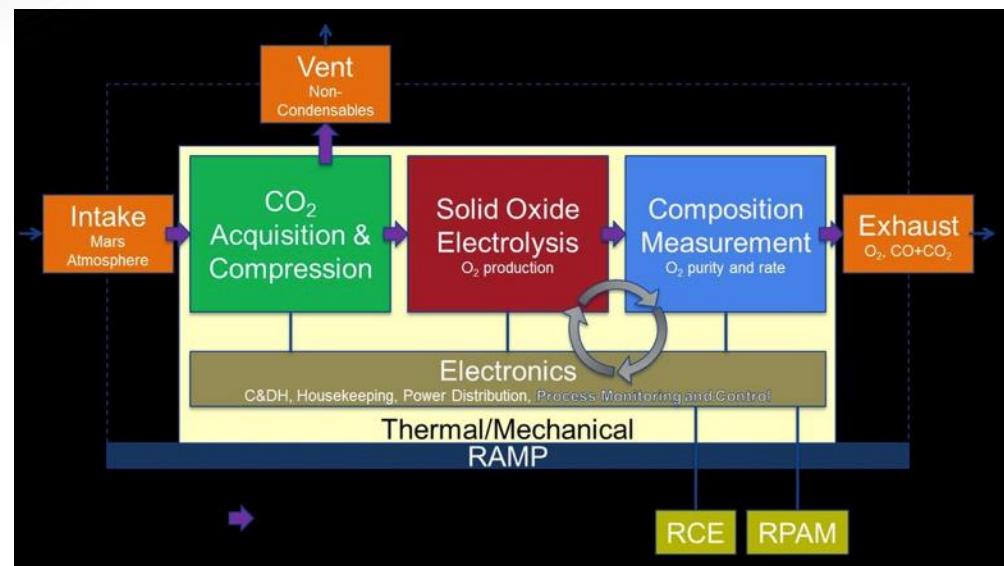
But...

- Usually have slower growth rates than heterotrophs
- Require photobioreactors

MOXIE: Mars Oxygen ISRU Experiment



Clostridium spp.



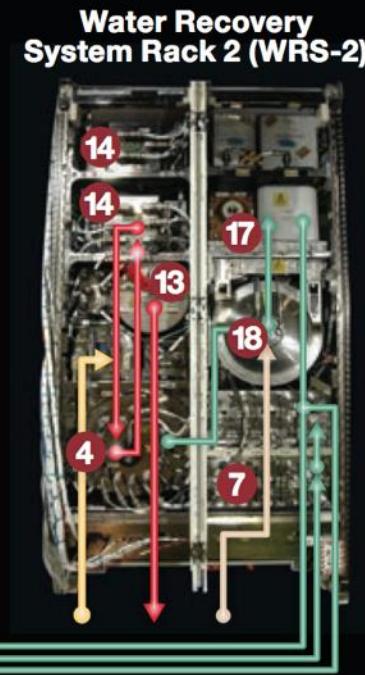
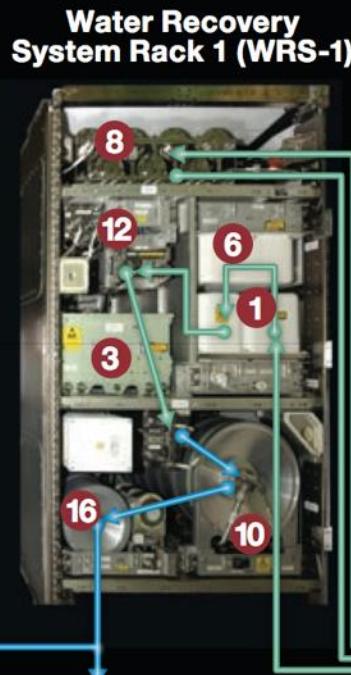
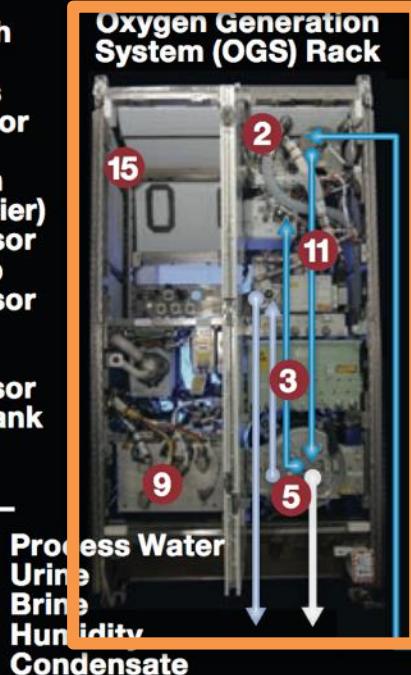
Picture Credits: NASA

Environmental Control and Life Support System

U.S. Regenerative Environmental Control and Life Support System (ECLSS)

1	Catalytic Reactor	12	Reactor Health Sensor
2	Deionizer Beds	13	Storage Tanks
3	Digital Controller	14	Urine Processor
4	Distillation Assembly	15	Pumps
5	Electrolysis Cell Stack	16	CO ₂ Reduction System (Sabatier)
6	Gas Separator	17	Water Processor
7	Multifiltration Beds	18	Delivery Pump
8	Particulate Filter		Water Processor Pump & Separator
9	Power Supply		Water Processor Wastewater Tank
10	Product Water Tank		
11	Pumps & Valves		

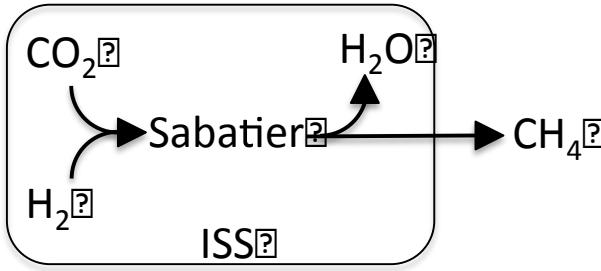
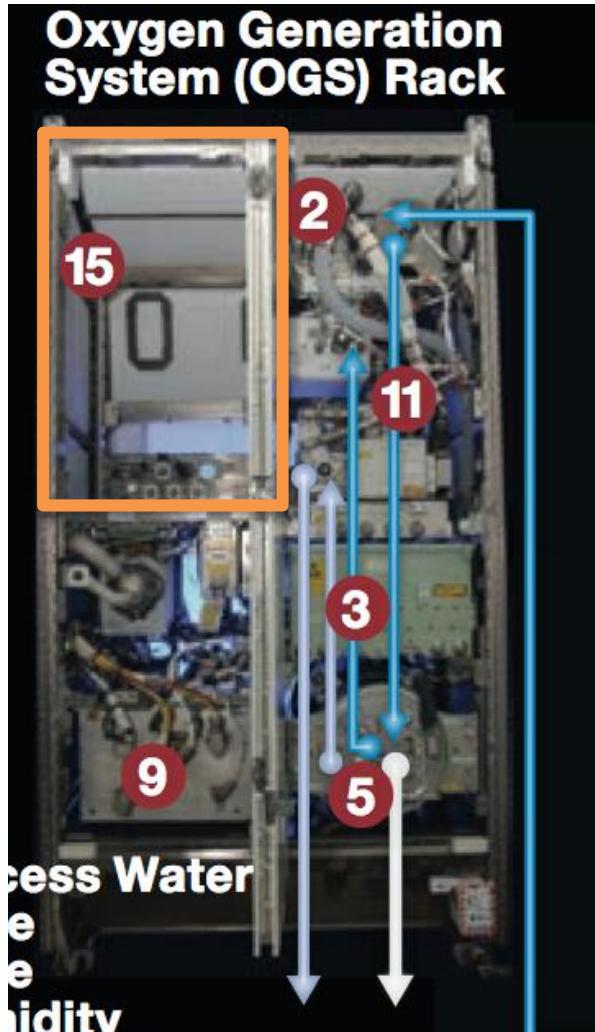
— = Oxygen
— = Hydrogen
— = (vented overboard)
— = Potable Water



Credits: NASA

“Reference Guide to the International Space Station”

Methane production by Sabatier



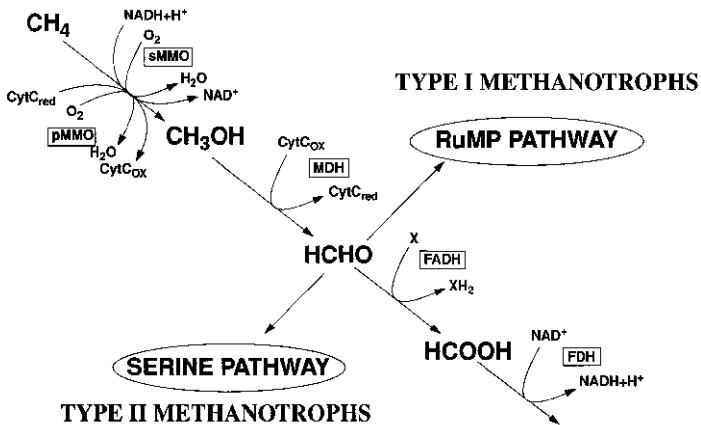
- Reacts CO_2 and H_2 to form CH_4 and H_2O
- H_2O is recycled
- CH_4 vented to space
- Operational since 2010

Astronaut Doug Wheelock
installing Sabatier system on ISS

“Waste” CH_4 could be fed to methanotrophs in a microbial manufacturing scheme.

Why Not Bacterial Methanotrophs?

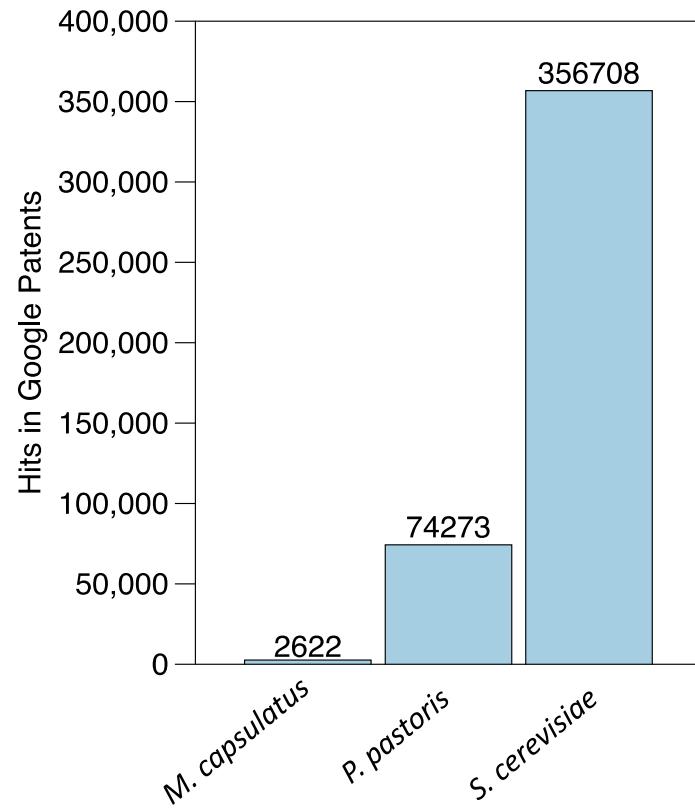
M. capsulatus



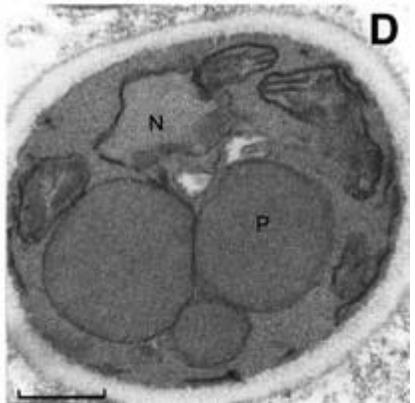
J. Bacteriol. October 2003
vol. 185 no. 19 5755-
5764

Microbiol. Mol. Biol. Rev. June
1996 vol. 60 no. 2 439-471

- NASA STTR to Mango Materials (Small Technology Transfer Innovation Research)
- “A Novel, Membrane-Based Bioreactor Design to Enable a Closed-Loop System on Earth and Beyond”
- “...a membrane bioreactor system to produce a biopolymer from methane gas...will enable bacterial growth and biopolymer production to occur in microgravity environments...”

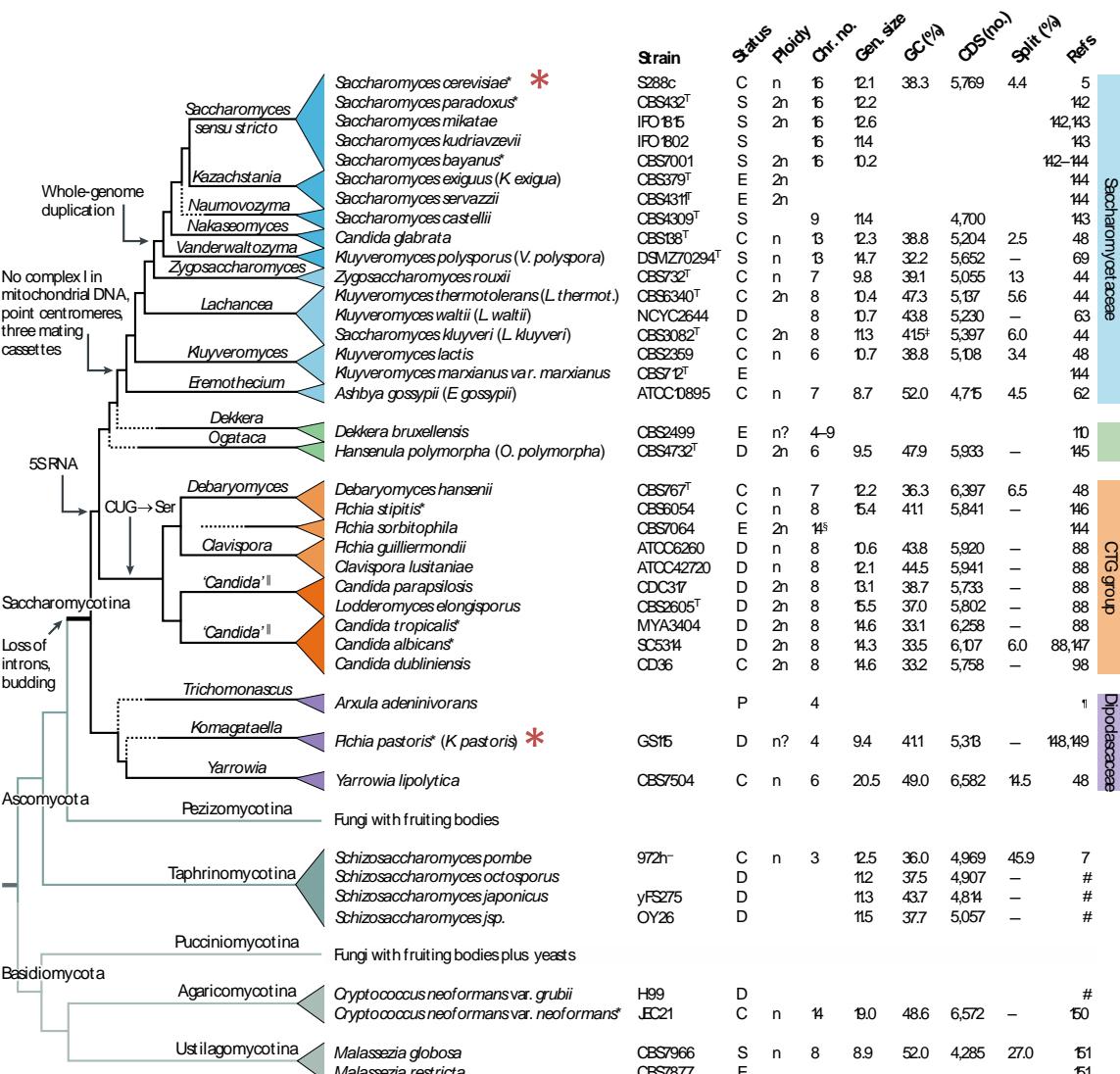


Pichia pastoris (*Komagataella phaffii*)



Ida J. van der Klei et al. EMBO J. 1998;17:3608-3618

- Diverged from *S. cerevisiae* ~200 MYA
- Methylotrophic yeast
- Widely used as a protein production host
- Capable of producing reduced and glycosylated proteins
- Grows to very high cell density (optical densities up to 630, Dr. Julia Cino, New Brunswick Scientific)



Dujon, 2010, Nature Reviews Genetics

Why *Pichia pastoris*?

1. Established synthetic biology platform

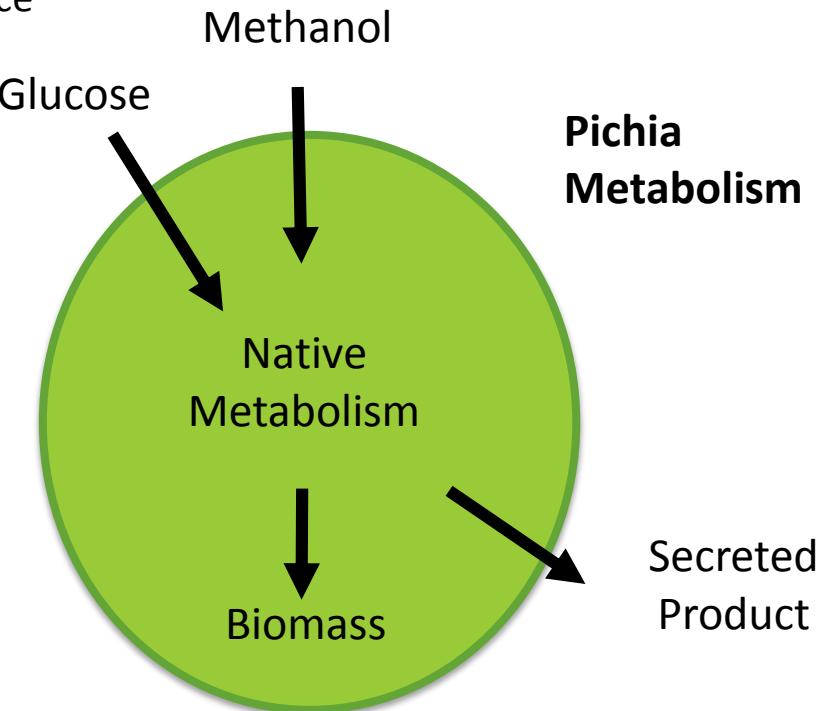
Used to produce Trypsin, murine TNF α , and FDA approved drugs Kalbitor (60 amino acid peptide to treat hereditary angioedema) and Jetrea (proteolytic enzyme to treat symptomatic vitreomacular adhesion)

2. Methylotrophic yeast

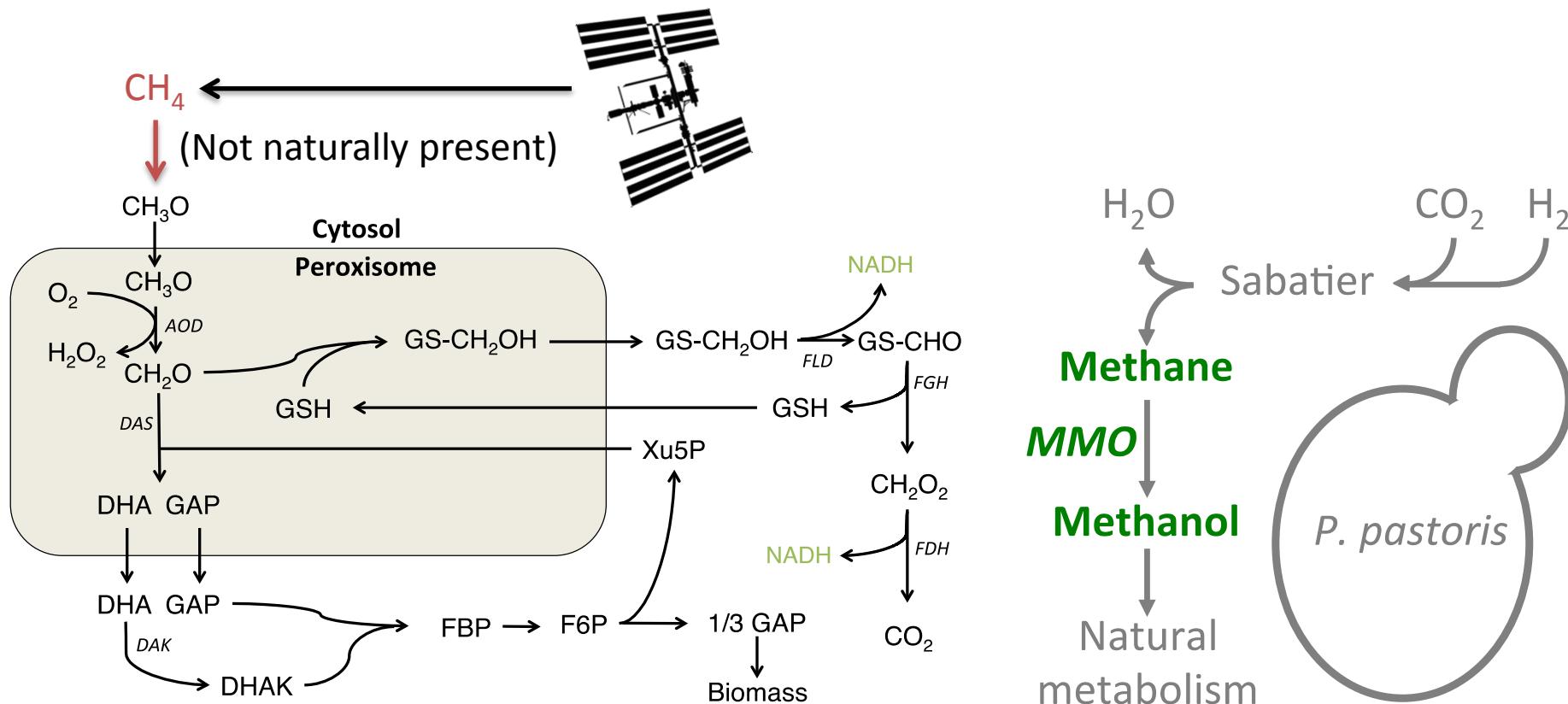
Can utilize methanol (CH_3OH) as a carbon source



Dennis Kunkel Microscopy, 2017,
Science

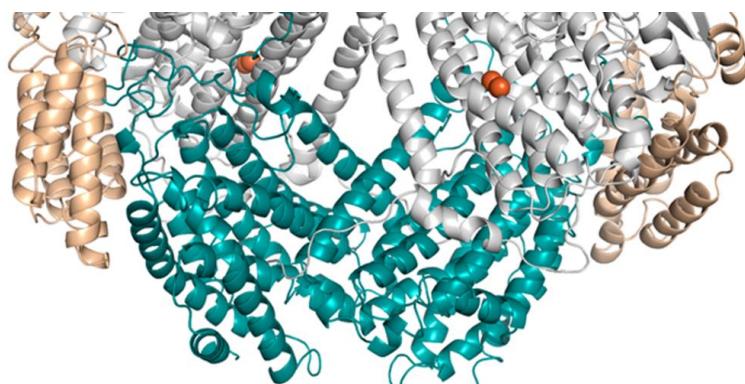
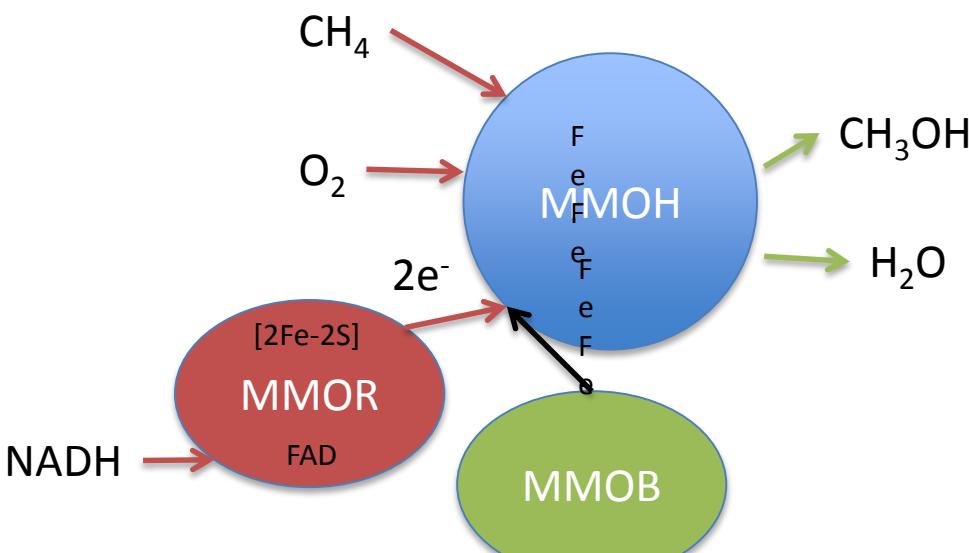


Porting methane metabolism to *P. pastoris*



Methanol (CH₃O) is oxidized to formaldehyde (CH₂O) by alcohol oxidase (AOD). Formaldehyde can either be oxidized to CO₂ through the successive action of formaldehyde dehydrogenase (FLD), S-formylglutathione hydrolase (FGH), and formate dehydrogenase (FDH), or appended to xylulose-5-phosphate and assimilated into biomass through a pathway involving dihydroxyacetone synthase (DAS) and dihydroxyacetone kinase (DAK).

Porting Soluble Methane Monooxygenase to *Pichia*



Sirajuddin and Rosenzweig. 2015.

1. Hydroxylase: MMOH

1. alpha
2. beta
3. gamma

Oxidizes methane hydroxylates methane to methanol

2. Reductase: MMOR

Oxidizes NADH and transfers electrons to MMOH

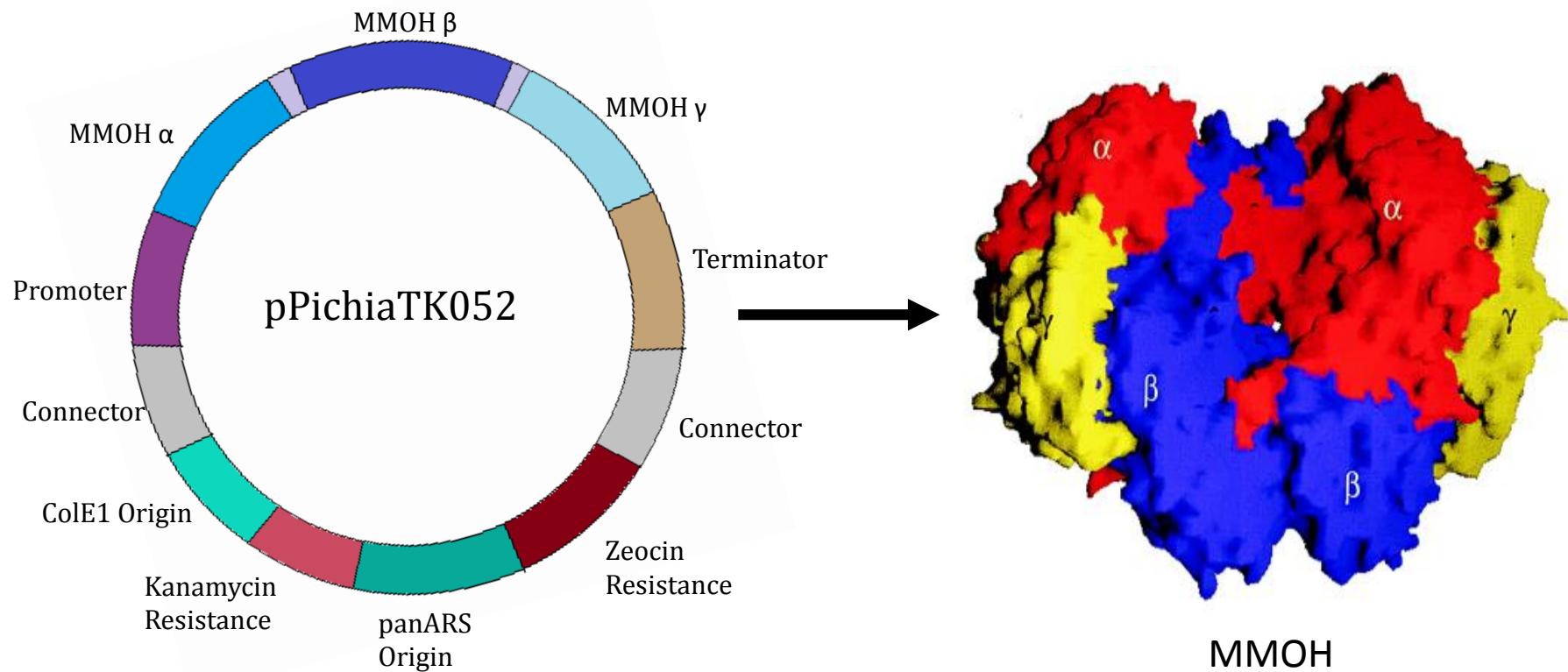
1. Regulatory: MMOB

Binds to same site as MMOR. May help drive cycle.

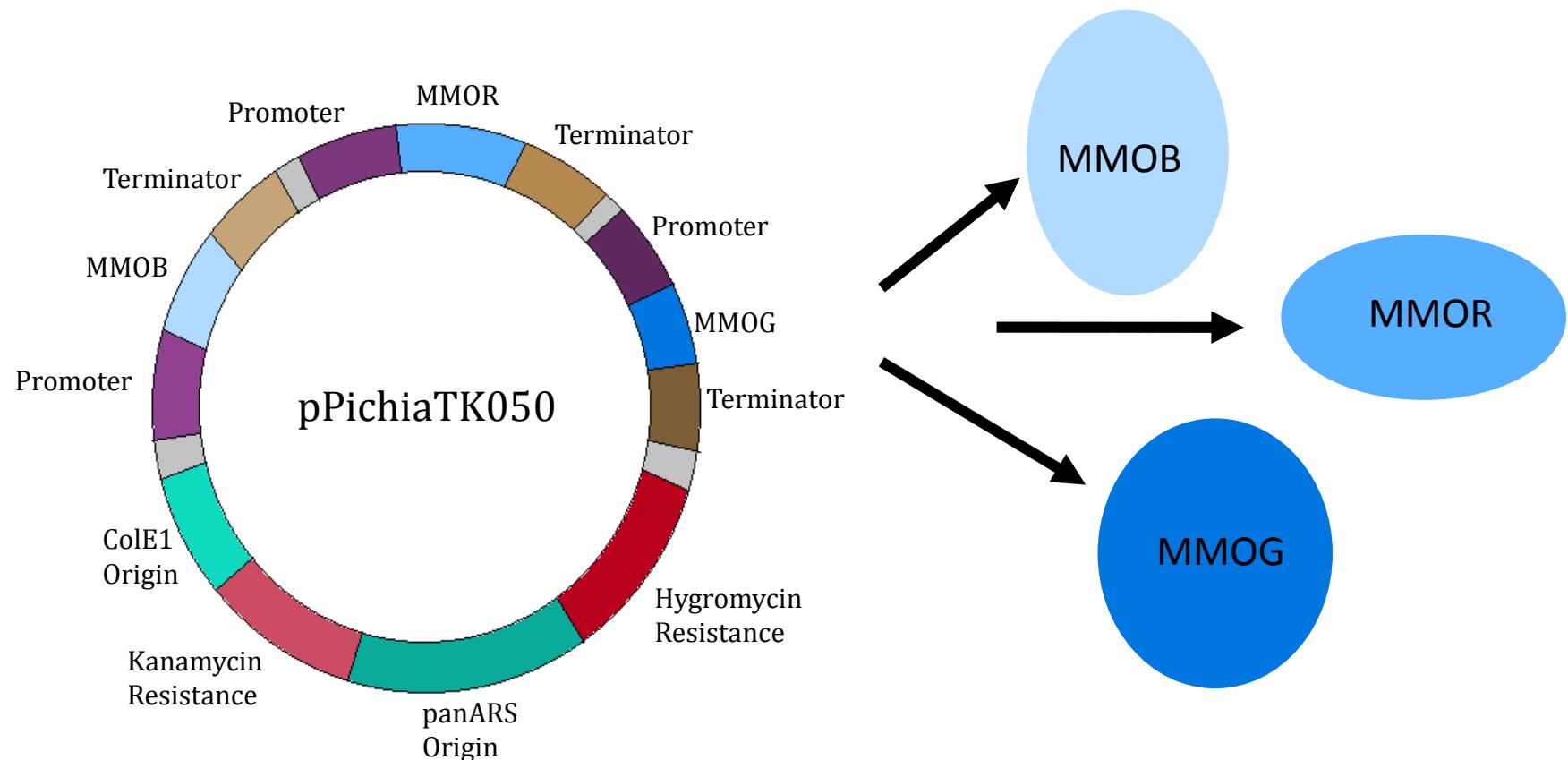
2. Assembly chaperone: MMOG

Could help in assembly of MMOH

Plasmid I: Single transcript expression of MMOH



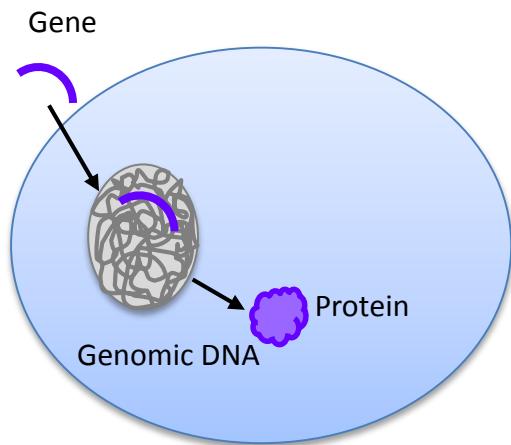
Plasmid II: Expression of accessory proteins



Introducing MMO genes to *Pichia*

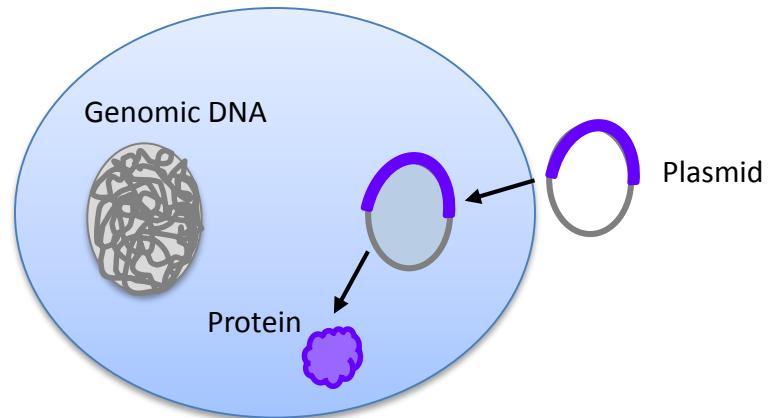
Genome Editing

- Insert gene into organismal genome
- CRISPR
- Homologous Recombination



Plasmid Transformation

- Gene inserted as a small independently replicating circle of DNA



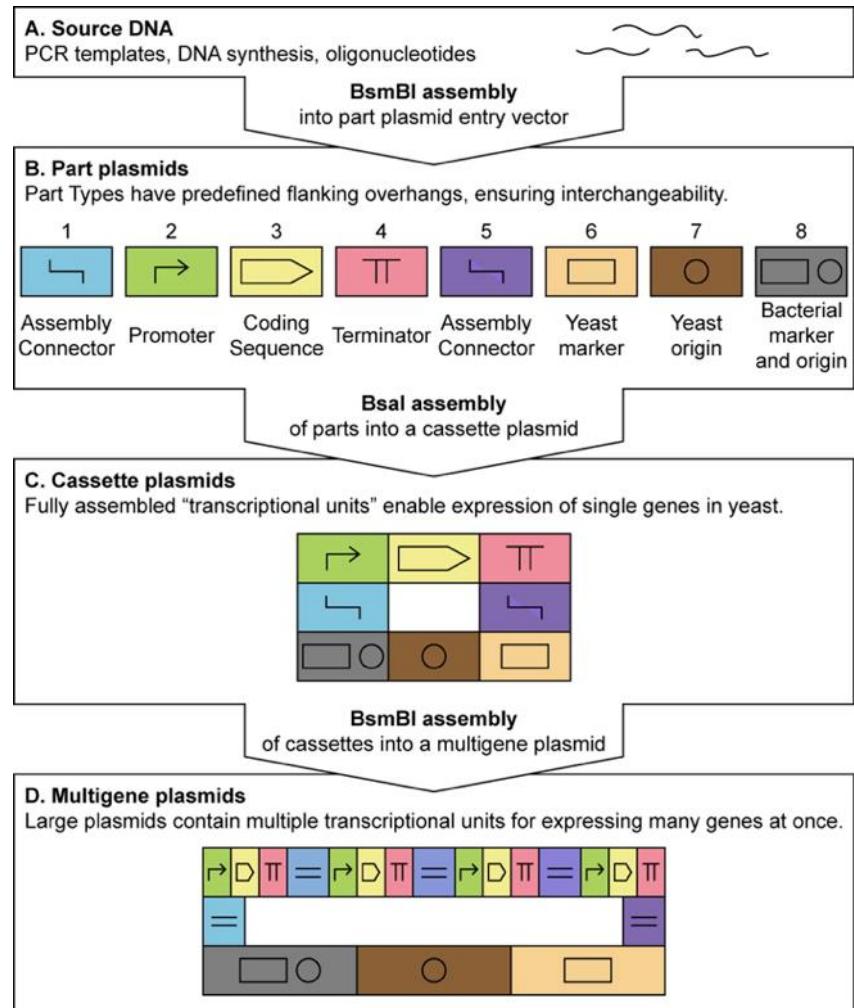
Developing *Pichia* tools

Pichia tools are limited compared to *S. cerevisiae*

Plasmid systems have only recently been developed

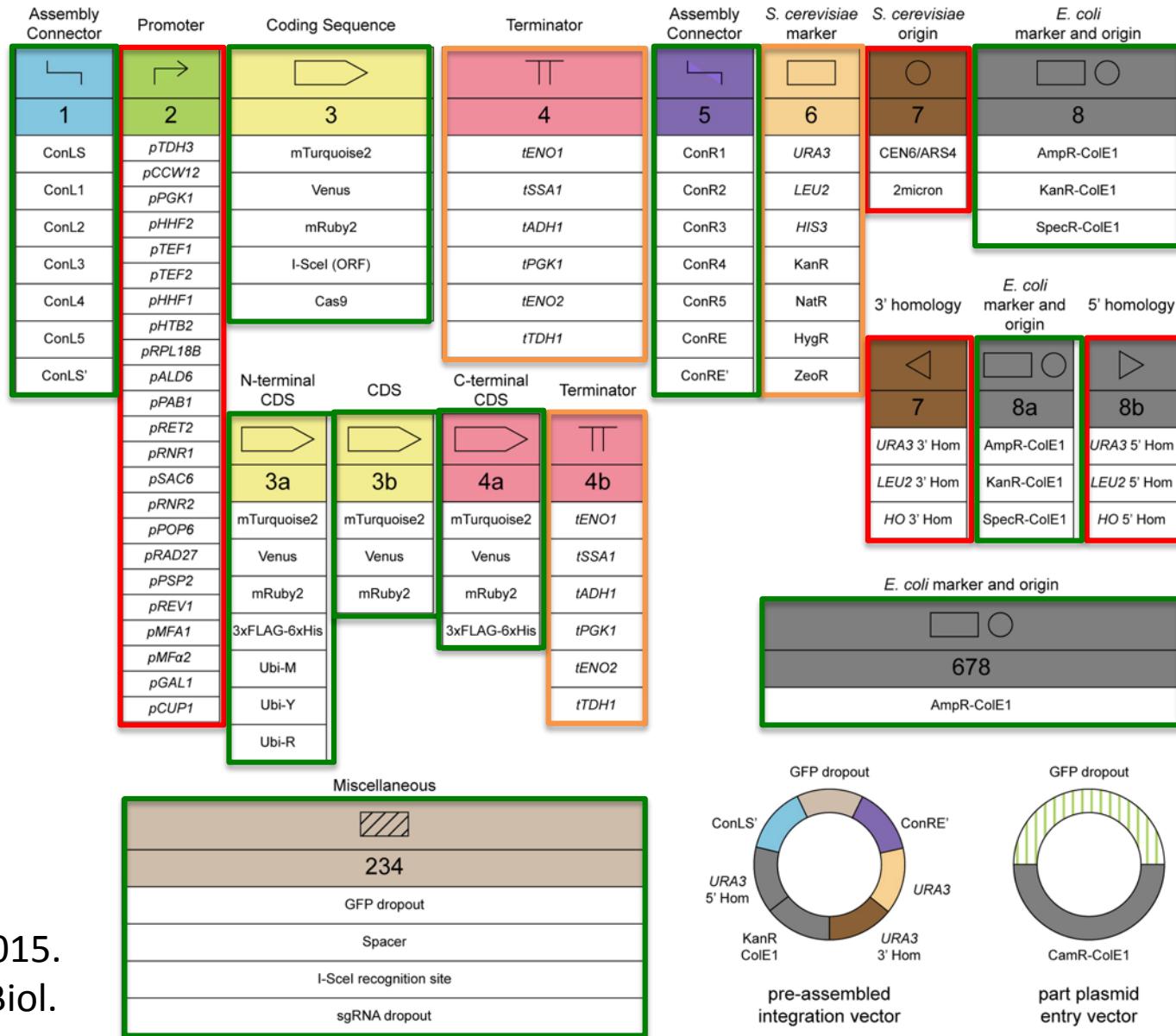
John Dueber at Berkeley has developed a “*S. cerevisiae* Toolkit”

This could be adapted for use in *Pichia*



Lee et al. 2015. ACS Synth Biol.

Building a *Pichia* toolkit

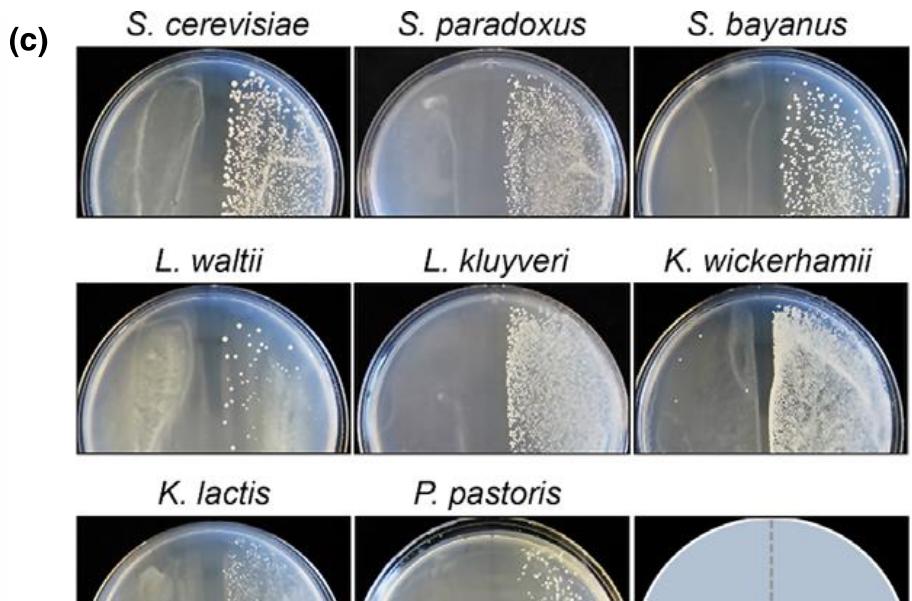
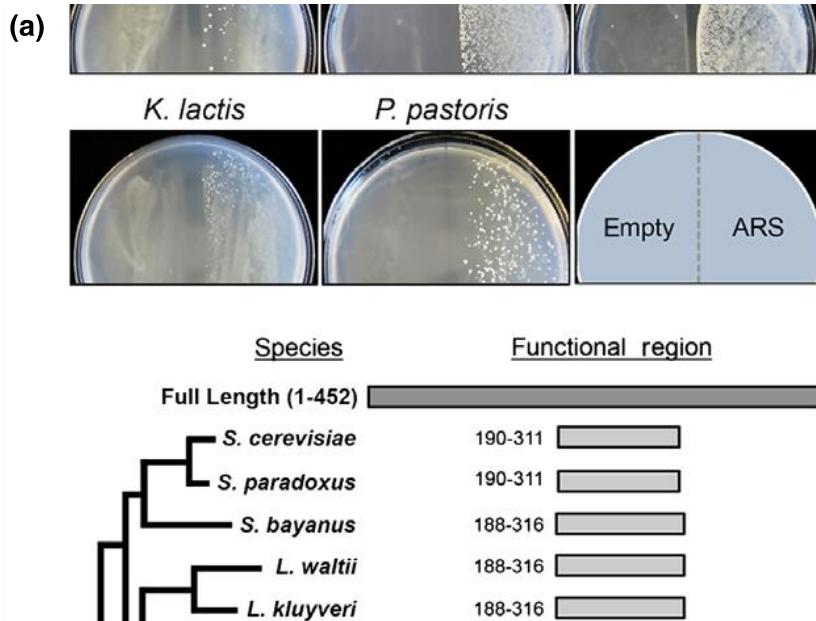


Origin of replication: panARS

An autonomously replicating sequence for use in a wide range of budding yeasts

Ivan Liachko & Maitreya J. Dunham

Department of Genome Sciences, University of Washington, Seattle, WA, USA

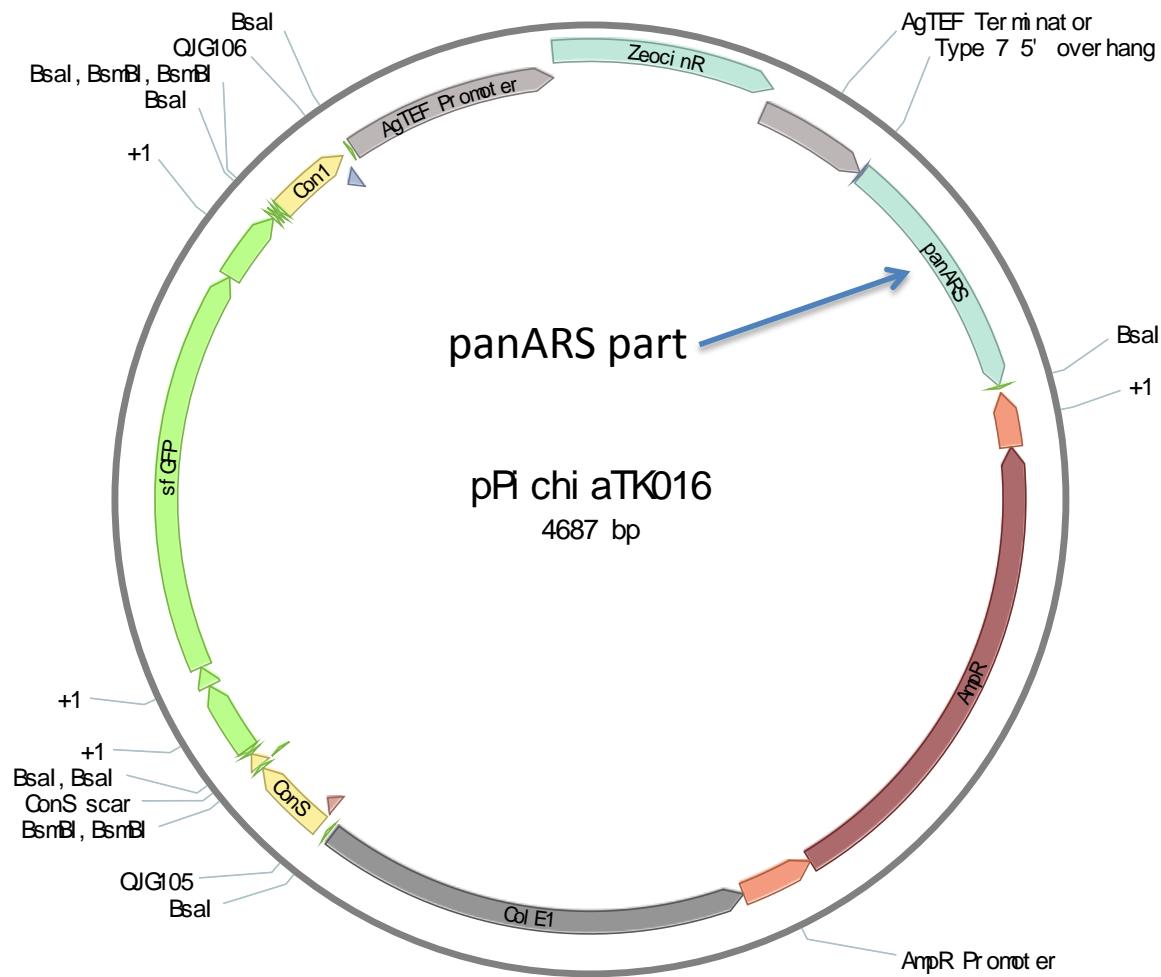


Plasmid system for *Pichia*

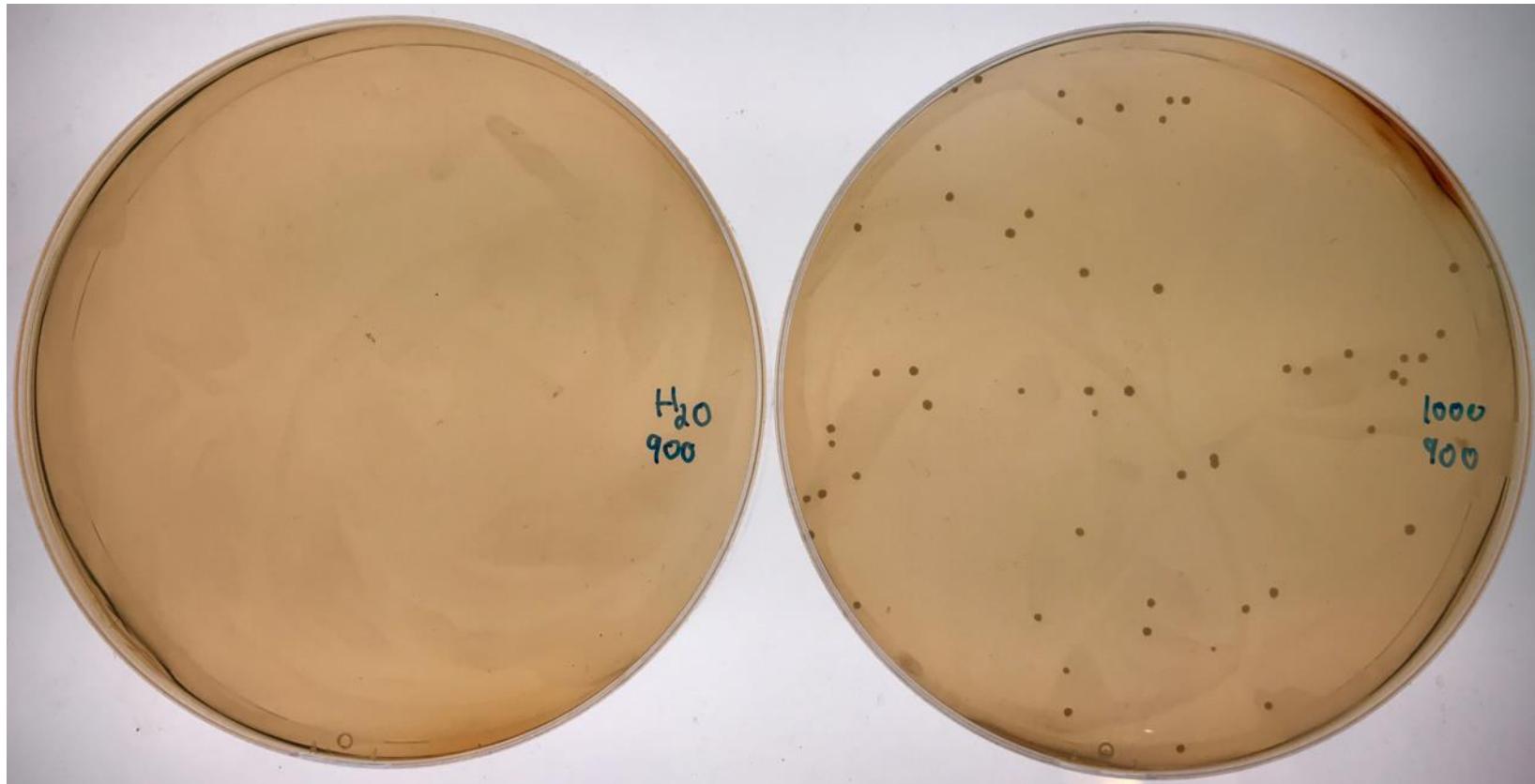
Adapted plasmid system from John Dueber by incorporating recently identified “panARS” sequence to allow replication of plasmid in *Pichia*.

Most other parts should be reusable.

Constructed empty vector that should impart resistance to Zeocin.



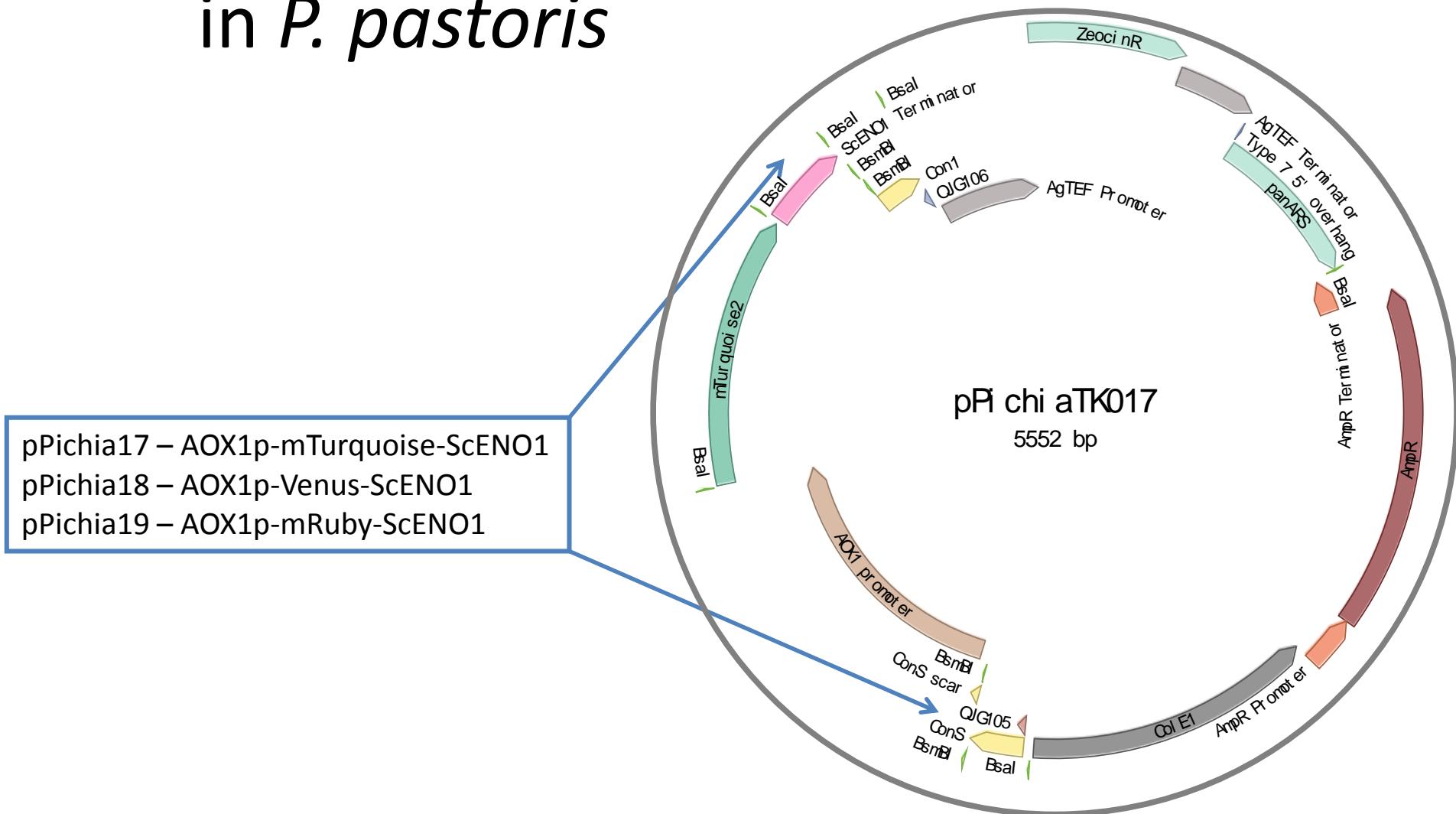
Plasmid system for *Pichia*



Water

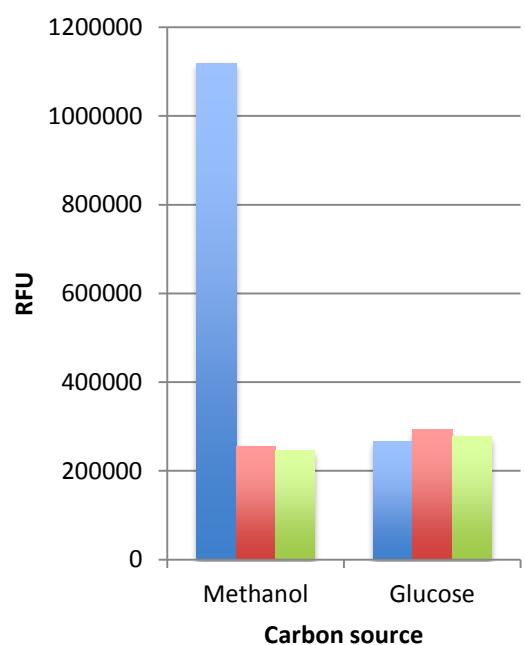
1 μ g pTK16

Plasmid demonstration in *P. pastoris*

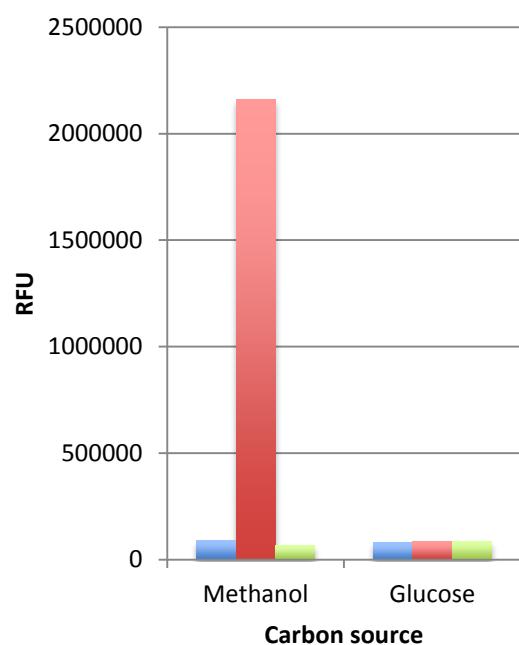


Plasmid demonstration in *P. pastoris*

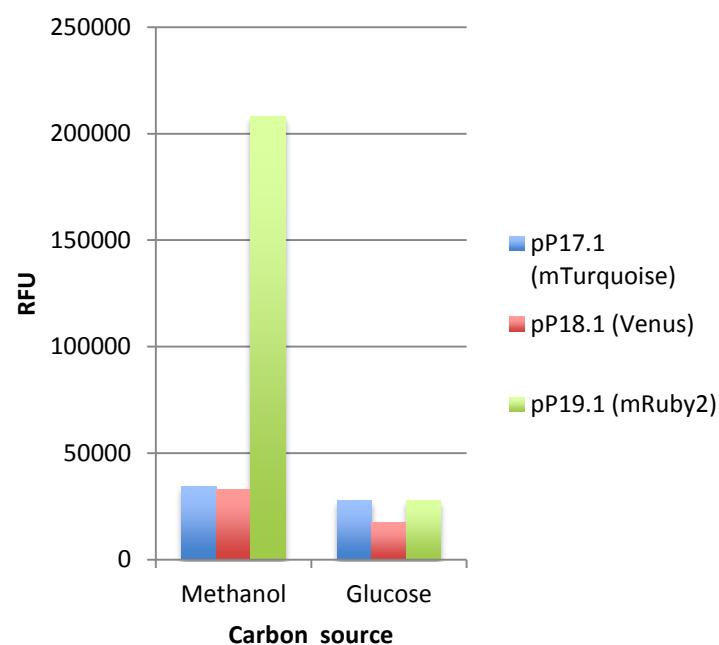
mTurquoise2



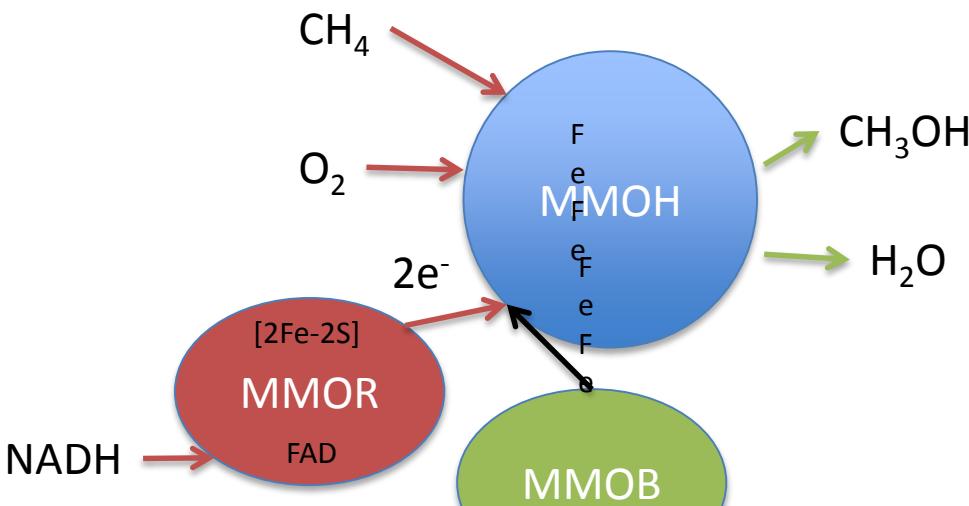
Venus



mRuby2



Porting Soluble Methane Monooxygenase to *Pichia*



1. Hydroxylase: MMOH

1. alpha
2. beta
3. gamma

Oxidizes methane hydroxylates methane to methanol

2. Reductase: MMOR

Oxidizes NADH and transfers electrons to MMOH

1. Regulatory: MMOB

Binds to same site as MMOR. May help drive cycle.

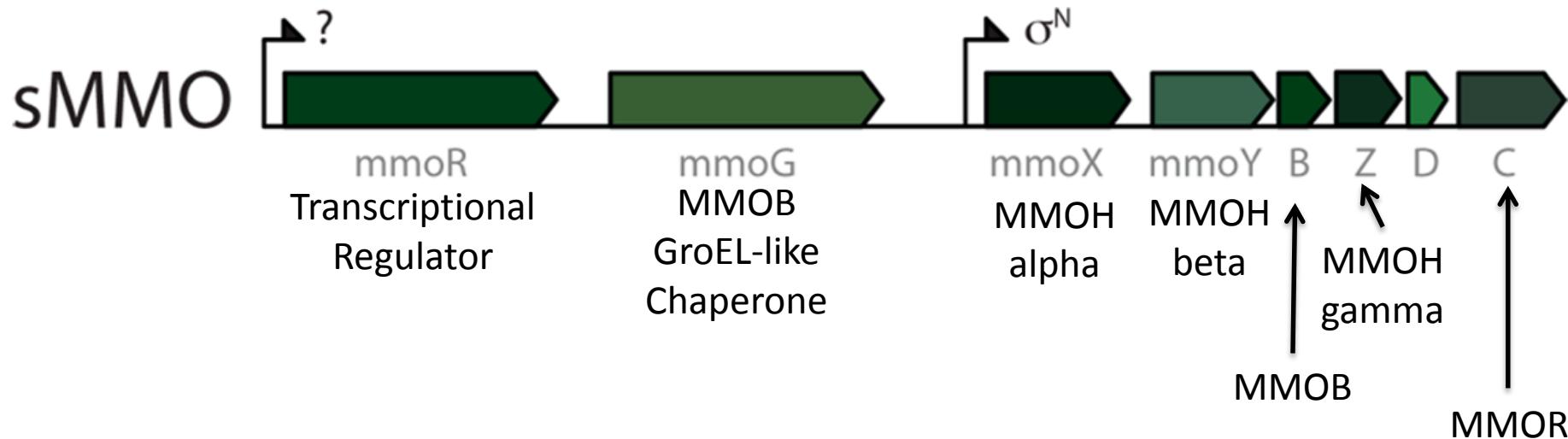
2. Assembly chaperone: MMOG

Could help in assembly of MMOH



Sirajuddin and Rosenzweig. 2015.

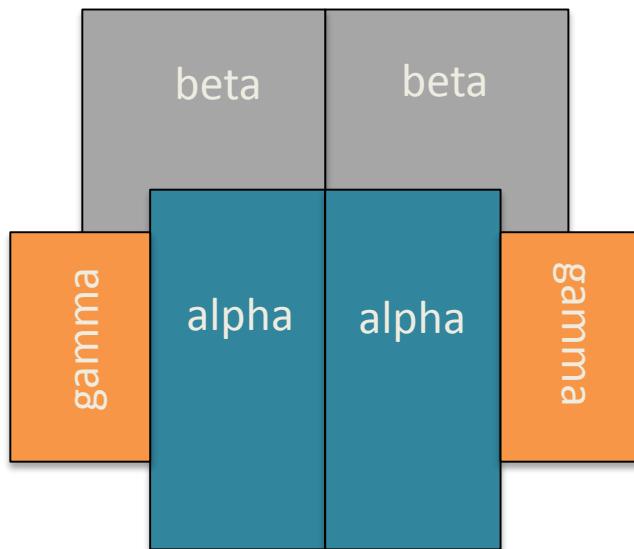
MMO components are expressed as an operon



- Most studies are sMMO from *Methylococcus capsulatus* (Bath) or *Methylosinus trichosporium* OB3b
- MMO from *M. trichosporium* OB3b has a higher turnover number (3.5 vs. $0.2 - 1.0 \text{ s}^{-1}$)

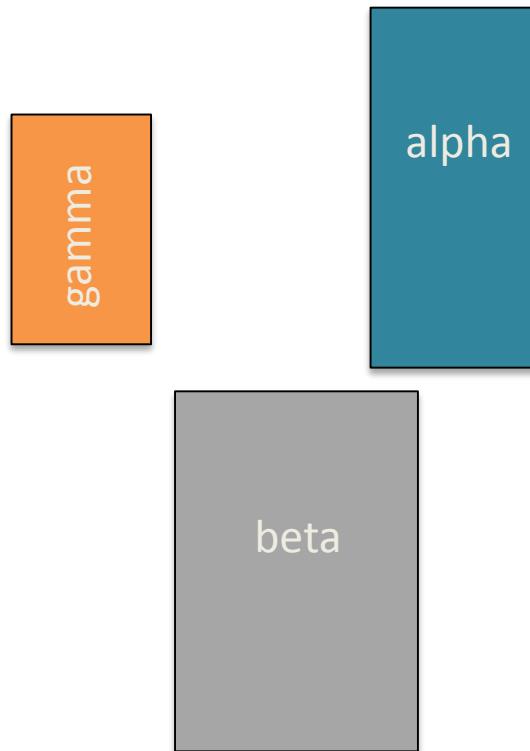
Proper assembly of MMOH is challenging

Assembled Complex



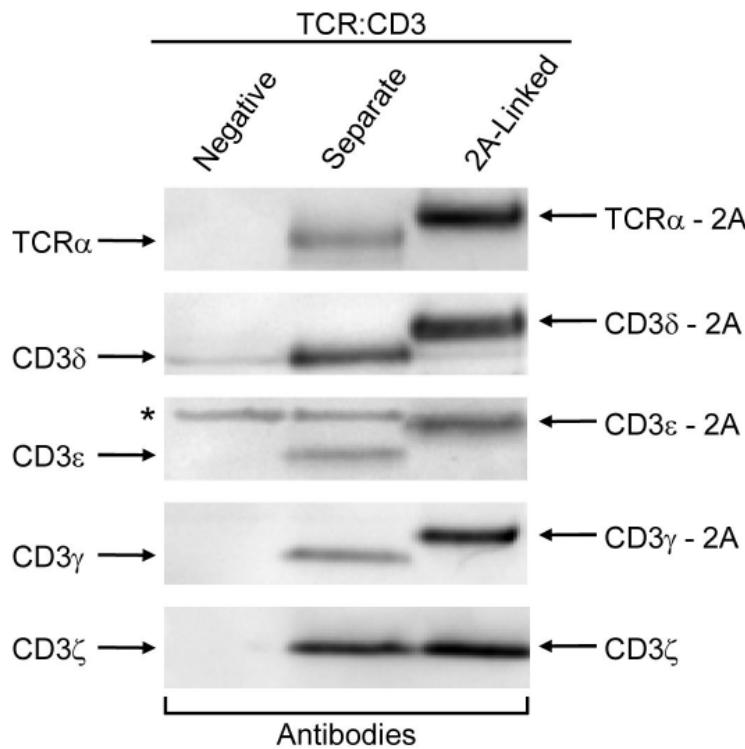
Stable. Active complexes are readily purified from *M. capsulatus*.

Pre-assembled Components



Unknown stability in *Pichia*. Assumed to be unstable.

Type 2a peptides balance component stoichiometry



VKQTLNFCLLKLAGDVESNP GP



Ribosome skips this peptide bond

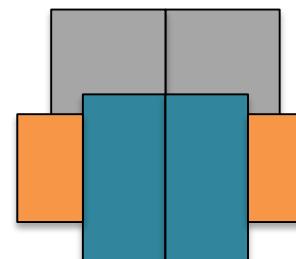
MMOH

Alpha---2a---Beta---2a---Gamma

Alpha-2a

Beta-2a

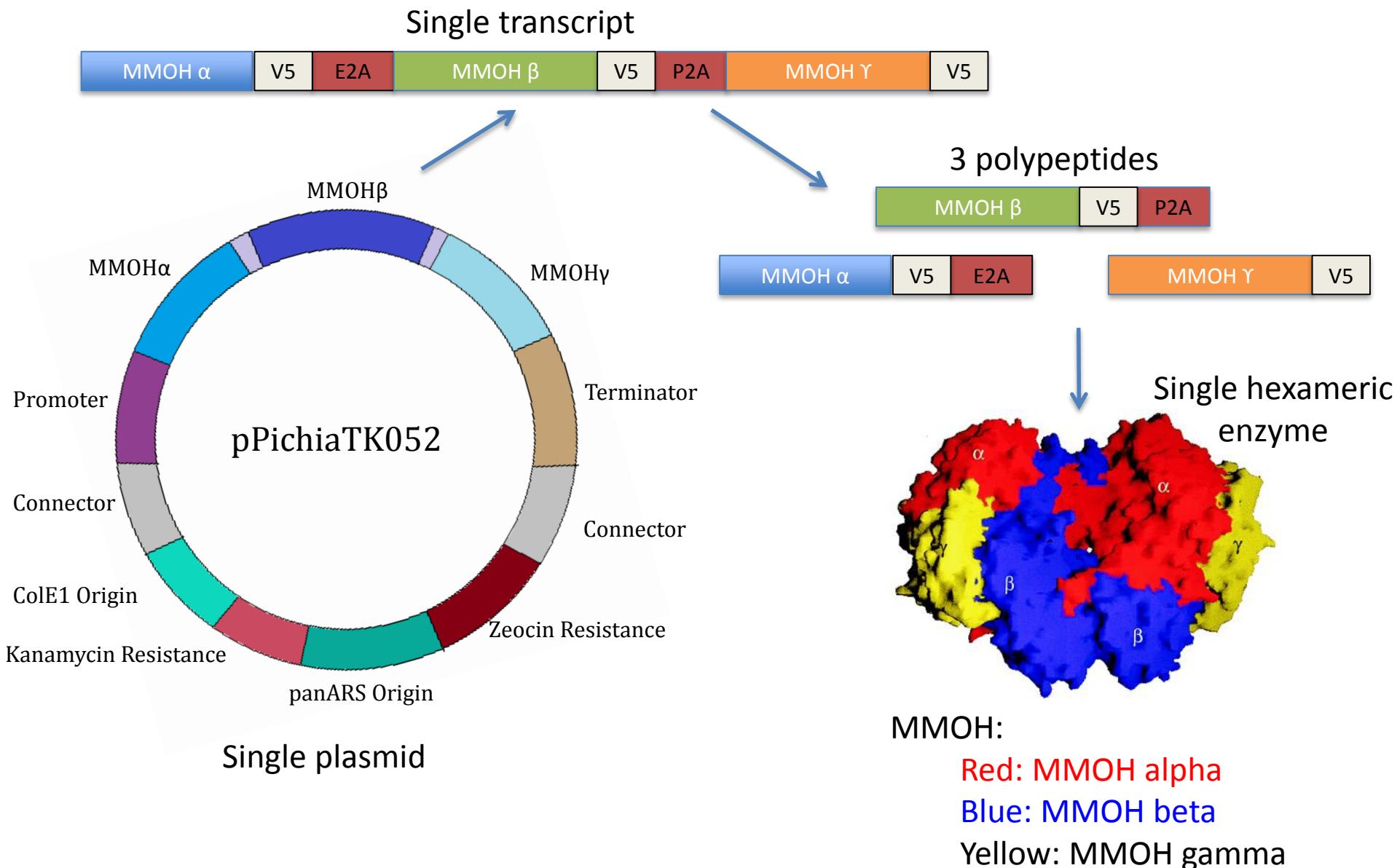
Gamma



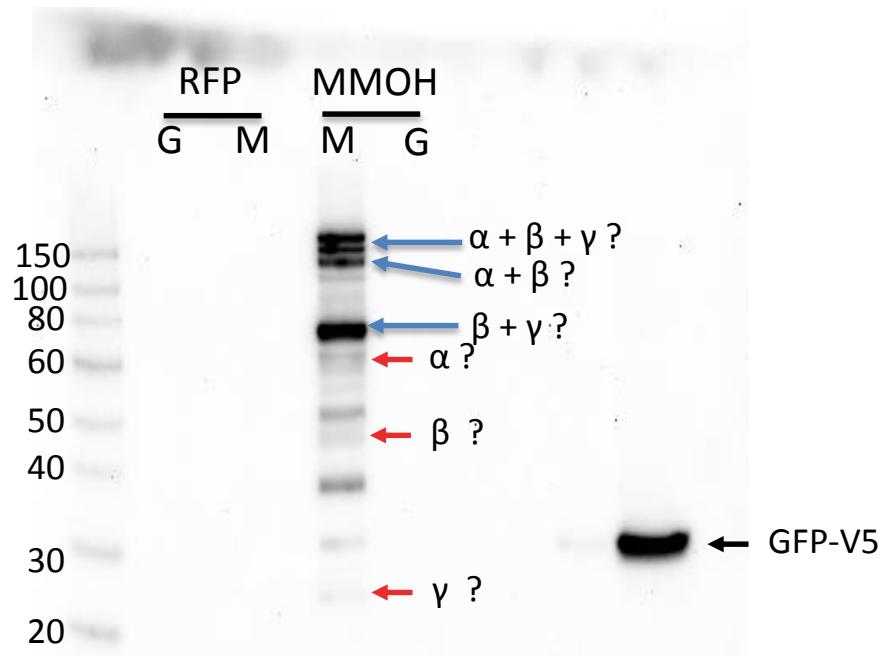
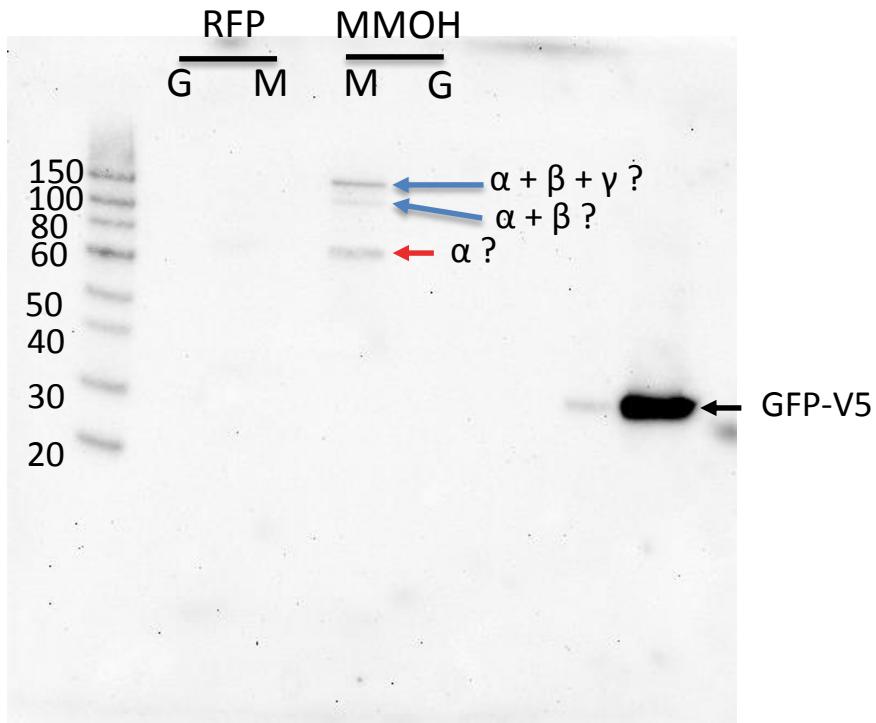
Stoichiometric production of complex components

Szymczak AL et al. *Nat Biotechnol*
2004; **22**: 589–594; Epub 2004, April
4.

Expressing MMOH from single transcript

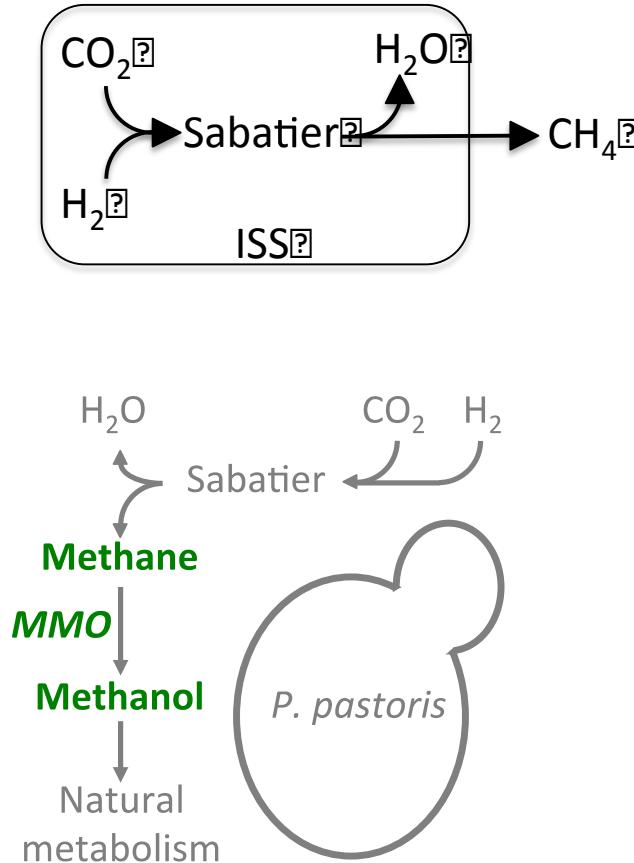


Western blot to detect MMOH subunits



Protein	MW (kDa)
MMOH α	63.5
MMOH β	48.4
MMOH γ	21
MMOH $\alpha + \beta + \gamma$	132.9
MMOH $\alpha + \beta$	111.9
MMOH $\beta + \gamma$	69.4

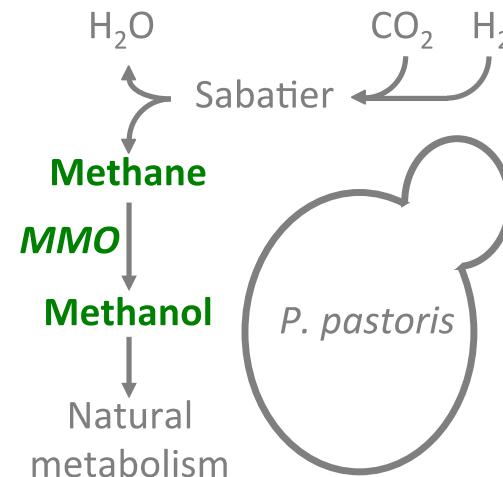
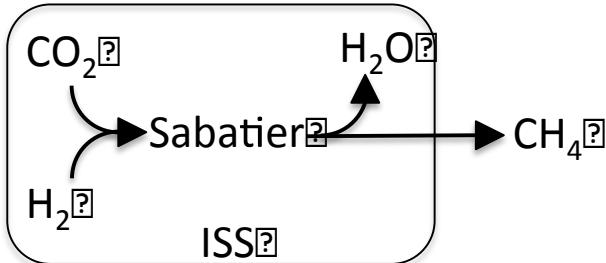
Porting sMMO to *Pichia*: Current status



Status

	Designed	Built	Tested
MMOH			<i>n.a.</i>
MMOB			<i>n.a.</i>
MMOR			<i>n.a.</i>
MMOG			<i>n.a.</i>
MMOH plasmid			
MMOR/B/G plasmid			

Potential of Biotechnology in Space



Credits: NASA

Funding

Funding

NASA Space Technology Mission Directorate Center Innovation Fund
NASA Advanced Exploration Systems

Team

Dr. John A. Hogan (NASA)

Dr. Asif Rahman (U. of New Mexico)

Dr. Aditya Hindupur (Wyle Labs)



NASA Ames Team Members

Dr. Michael Dougherty (Wyle Labs)
Natalie Ball (Wyle Labs)
Dr. Hiromi Kagawa (SETI)



Dr. John Hogan



Samantha Fleury & Lily Neff

WASHU Collaborators

Fuzhong Zhang Chris Bowen

Cameron Sargent Sarah Rommelfanger